



## Regioselective iodination of phenol and analogues using *N*-iodosuccinimide and *p*-toluenesulfonic acid

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### ABSTRACT

Mild and highly regioselective monoiodination of phenol and analogues is achieved in high to excellent yields at room temperature with a combination of stoichiometric *p*-toluenesulfonic acid and *N*-iodosuccinimide.

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Iodination of phenols furnishes important synthetic intermediates<sup>1</sup> which are useful in a variety of palladium-catalysed and copper(I)-assisted reactions, such as the Heck, Stille and the Ullmann reactions.<sup>2</sup> Iodophenol compounds are also constituents of various naturally occurring biologically active compounds, especially those of marine sponge metabolites.<sup>3</sup> Numerous methods exist for direct iodination of phenols which include NaClO<sub>2</sub>-NaI-HCl,<sup>4a</sup> ICl,<sup>4b</sup> I<sub>2</sub>-tetrabutylammonium peroxydisulfate,<sup>4c</sup> bis(*sym*-collidine)iodine(I) hexafluorophosphate,<sup>4d</sup> benzyltrimethylammonium dichloroiodate,<sup>4e</sup> chloramine T-NaI<sup>4f</sup> and NaI-*tert*-butyl hypochlorite.<sup>4g</sup> Previous regioselective phenol iodinations employed various combinations of reagents, that include I<sub>2</sub>-cerium ammonium nitrate,<sup>5a</sup> I<sub>2</sub>-β-cyclodextrin,<sup>5b</sup> TIOAc-I<sub>2</sub><sup>5c</sup> and KI-benzyltriphenylphosphonium peroxymonosulfate.<sup>5d</sup> While several methods to achieve *ortho*-iodination have been established,<sup>5a–c</sup> high yielding *para*-selective iodination of phenol and analogues remains a challenge.

As part of our continuing efforts to synthesise L-thyronine analogues, via L-tyrosine and *para*-iodophenol derivative copper(I)-assisted coupling reactions, we were obliged to seek a mild, facile and highly regioselective methodology for the synthesis of halogenated analogues of *p*-iodophenol, such as 6-bromo-2-chloro-4-iodophenol. A potential source of iodonium ions, other than molecular iodine itself, is *N*-iodosuccinimide (NIS). The use of NIS in aromatic electrophilic iodination has been amply demonstrated in deactivated and activated aromatic systems using various NIS-activating co-reagents, such as CF<sub>3</sub>SO<sub>3</sub>H (as solvent for deactivated systems),<sup>6a</sup> catalytic hydroxy(tosyloxy)iodobenzene (HTIB),<sup>6b</sup> catalytic *p*-toluenesulfonic acid (*p*TsOH),<sup>6c</sup> CH<sub>3</sub>CN (as solvent for the activated systems)<sup>6d</sup> and catalytic CF<sub>3</sub>CO<sub>2</sub>H.<sup>6e</sup> Herein the combination of *p*TsOH and NIS, previously demonstrated in

the iodination of polyalkylbenzenes,<sup>6c</sup> and recently in the syntheses of 3-iodo and 3-halo-5-iodo analogues of *N*-acetyl-L-tyrosine,<sup>6f</sup> was found to give, at ambient temperature, highly regioselective monoiodination of phenol and its analogues in high to excellent yields.

Using 1 equiv of *p*TsOH and NIS in acetonitrile, phenol (**1**) was converted, after 14 h of stirring at room temperature, to 4-iodophenol (**1a**) in 95% yield (Table 1, entry **1b**). Acids other than *p*TsOH were also investigated for their promoting effect on the selectivity of *para*-iodination of phenol. With 1 equiv of CH<sub>3</sub>CO<sub>2</sub>H, the yield and selectivity of **1a** were 14% and 16%, respectively. With 1 equiv of H<sub>3</sub>PO<sub>4</sub>, HCl or H<sub>2</sub>SO<sub>4</sub>, the yields and selectivities of **1a** were 42% and 44%, 61% and 81%, and 77% and 82%, respectively. In the absence of *p*TsOH or any other acids (Table 1, entry **1a**), the selectivity and yield of **1a** were poor and the reaction was accompanied by products of di and triiodination at positions 2,4 and 2,4,6, respectively. For other *ortho* and *para*-substituted phenols, iodination without the promoting acid also afforded diiodinated by-products in low to moderate yields. The promoting-effect of *p*TsOH on the selectivity of iodination, as demonstrated by the change in sequence of *p*TsOH addition to the acetonitrile-substrate solution (adding last after NIS), showed that all, except for the iodination of **6**, gave poor yields of the desired products, **1a–11a** (Table 1, entries **1c–11c**). We speculate that the role of *p*TsOH in the reaction is to suppress the formation of the phenolate anion and as a result, diminish iodination via the phenolic anion.

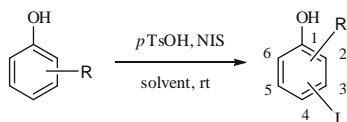
The most effective amount of *p*TsOH to promote optimum yield and regioselectivity was found to be 1 equiv. At 0.05 equiv of *p*TsOH, the yield and selectivity of **1a** were 69% and 89%, respectively. Increasing the amount of *p*TsOH to 0.1, 0.3 and 0.5 equiv did not improve the yields and selectivities of **1a**, which remained at 70% and 87–89%, respectively.

The best solvent for effecting regioselective iodination of phenols and analogues was acetonitrile, as was also shown in the

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**Table 1**Effects of *p*-toluenesulfonic acid, solvents and temperature on the monoiodination of phenol and analogues<sup>a</sup>R = H, 2,3-C<sub>4</sub>H<sub>4</sub>, 3,4-C<sub>4</sub>H<sub>4</sub>, 2-OCH<sub>3</sub>, 2-Br, 2-Cl, 4-Cl, 2-CH<sub>3</sub>, 4-CH<sub>3</sub>, 2-COCH<sub>3</sub>, 2-CO<sub>2</sub>CH<sub>3</sub>

Entry	Substrate	Solvent	Product composition <sup>a</sup> (%)							Product	Isolated yield (%)	
			S.M.	2	4	6	2,4	2,6	4,6			2,4,6
<b>1a</b>		CH <sub>3</sub> CN <sup>b</sup>	29	3	26	—	20	0	—	22		95
<b>1b</b>		CH <sub>3</sub> CN <sup>a</sup>	0	4	<b>95</b>	—	1	0	—	0		
<b>1c</b>		CH <sub>3</sub> CN <sup>c</sup>	30	3	33	—	21	0	—	13		
<b>1d</b>		CH <sub>3</sub> CN <sup>a,d</sup>	8	6	86	—	0	0	—	0		
<b>1e</b>		CH <sub>3</sub> CN <sup>a,e</sup>	11	5	84	—	0	0	—	0		
<b>1f</b>		CH <sub>3</sub> OH	2	17	77	—	2	2	—	0		
<b>1g</b>		EtOAc	99	0	1	—	0	0	—	0		
<b>1h</b>		THF	2	16	78	—	2	2	—	0		
<b>1i</b>		1,4-Dioxane	20	22	58	—	Trace	Trace	—	0		
<b>1k</b>		CHCl <sub>3</sub>	0	13	<b>86</b>	—	1	0	—	0		
<b>2a</b>		CH <sub>3</sub> CN <sup>b</sup>	81	17	—	—	—	2	—	—		85
<b>2b</b>		CH <sub>3</sub> CN <sup>a</sup>	5	<b>95</b>	—	—	—	0	—	—		
<b>2c</b>		CH <sub>3</sub> CN <sup>c</sup>	77	20	—	—	—	3	—	—		
<b>3a</b>		CH <sub>3</sub> CN <sup>b</sup>	69	—	6	18	—	—	7	—		89
<b>3b</b>		CH <sub>3</sub> CN <sup>a</sup>	0	—	<b>91</b>	Trace	—	—	8	—		
<b>3c</b>		CH <sub>3</sub> CN <sup>c</sup>	30	—	46	11	—	—	13	—		
<b>4a</b>		CH <sub>3</sub> CN <sup>b</sup>	100	—	0	0	—	—	0	—		66
<b>4b</b>		CH <sub>3</sub> CN <sup>a</sup>	7	—	<b>87</b>	6	—	—	0	—		
<b>4c</b>		CH <sub>3</sub> CN <sup>c</sup>	100	—	0	0	—	—	0	—		
<b>5a</b>		CH <sub>3</sub> CN <sup>b</sup>	84	—	10	6	—	—	0	—		88
<b>5b</b>		CH <sub>3</sub> CN <sup>a</sup>	2	—	<b>95</b>	3	—	—	0	—		
<b>5c</b>		CH <sub>3</sub> CN <sup>c</sup>	4	—	92	4	—	—	0	—		
<b>6a</b>		CH <sub>3</sub> CN <sup>b</sup>	99	—	<1	0	—	—	0	—		73
<b>6b</b>		CH <sub>3</sub> CN <sup>a</sup>	23	—	<b>75</b>	2	—	—	0	—		
<b>6c</b>		CH <sub>3</sub> CN <sup>c</sup>	17	—	<b>80</b>	3	—	—	0	—		
<b>7a</b>		CH <sub>3</sub> CN <sup>b</sup>	36	—	16	4	—	—	44	—		78
<b>7b</b>		CH <sub>3</sub> CN <sup>a</sup>	14	—	<b>82</b>	4	—	—	0	—		
<b>7c</b>		CH <sub>3</sub> CN <sup>c</sup>	27	—	23	8	—	—	42	—		
<b>7d</b>		CHCl <sub>3</sub> <sup>a</sup>	6	—	<b>76</b>	18	—	—	0	—		
<b>7e</b>		CH <sub>3</sub> CN <sup>a</sup>	0	—	7	0	—	—	19 <sup>f</sup>	<b>74<sup>g</sup></b>		54
<b>8a</b>		CH <sub>3</sub> CN <sup>b</sup>	66	30	—	—	—	4	—	—		51
<b>8b</b>		CH <sub>3</sub> CN <sup>a</sup>	9	<b>74</b>	—	—	—	17	—	—		
<b>8c</b>		CH <sub>3</sub> CN <sup>c</sup>	10	78	—	—	—	12	—	—		
<b>8d</b>		CHCl <sub>3</sub> <sup>a</sup>	10	<b>83</b>	—	—	—	7	—	—		

(continued on next page)

Table 1 (continued)

Entry	Substrate	Solvent	Product composition <sup>a</sup> (%)							Product	Isolated yield (%)	
			S.M.	2	4	6	2,4	2,6	4,6			2,4,6
<b>9a</b>		CH <sub>3</sub> CN <sup>b</sup>	45	—	18	4	—	—	33	—		81
<b>9b</b>		CH <sub>3</sub> CN <sup>a</sup>	2	—	<b>9a</b>	4	—	—	0	—		
<b>9c</b>		CH <sub>3</sub> CN <sup>c</sup>	15	—	65	9	—	—	11	—		
<b>10a</b>		CH <sub>3</sub> CN <sup>b</sup>	17	83	—	—	—	—	—		93	
<b>10b</b>		CH <sub>3</sub> CN <sup>a</sup>	1	<b>99</b>	—	—	—	—	—			
<b>10c</b>		CH <sub>3</sub> CN <sup>c</sup>	1	99	—	—	—	—	—			
<b>11a</b>		CH <sub>3</sub> CN <sup>b</sup>	100	0	0	—	0	—	—		78	
<b>11b</b>		CH <sub>3</sub> CN <sup>a</sup>	16	3	<b>81</b>	—	0	—	—			
<b>11c</b>		CH <sub>3</sub> CN <sup>c</sup>	95	2	3	—	0	—	—			

<sup>a</sup> Reaction conditions: *p*TsOH (0.5 mmol) was added to a stirred solution (solvent = 10 ml) containing the substrate (0.5 mmol) at room temperature or 0 °C. After 5 min, 1 equiv of NIS was added and the mixture was stirred for 14 h. The reaction was monitored by GC/MS until completion and the product composition was determined by GC/MS. The major products were isolated by silica gel column chromatography (10–20% CH<sub>2</sub>Cl<sub>2</sub>/hexanes) and characterised by GC/MS, <sup>1</sup>H and <sup>13</sup>C NMR. The numbers in the product composition denote the positions of iodination relative to the OH (numbered 1) of the phenol and S.M. refers to the substrate.

<sup>b</sup> Without *p*TsOH.

<sup>c</sup> *p*TsOH (1 equiv) was added to the stirred solution (solvent = 10 ml) containing the substrate and NIS after a 15-min delay.

<sup>d</sup> The reaction mixture was stirred at room temperature for 8 h.

<sup>e</sup> Reaction mixture was stirred at 0 °C for 8 h.

<sup>f</sup> 2,4-Dibromo-6-chlorophenol (12%) and 4,6-diiodo-2-chlorophenol (7%) were obtained.

<sup>g</sup> Compound **7b**.

syntheses of 3-halo and 3-halo-5-iodo analogues of *N*-acetyl-L-tyrosine.<sup>6f</sup> In acetonitrile, monoiodination of **1** at room temperature afforded **1a** in 95% yield and selectivity (Table 1, entry **1b**). In methanol, the yield and the selectivity of **1a** were 77% and 79%, respectively (Table 1, entry **1f**). On the other hand, there was no significant reaction in EtOAc (Table 1, entry **1g**). In THF and 1,4-dioxane, iodination of **1** gave moderate yields and selectivities of **1a** (Table 1, entries **1h** and **1i**). Chloroform was found to be an effective solvent for regioselective iodination, even though the yield of *ortho* iodination (13%) in CHCl<sub>3</sub> was higher compared to that obtained in acetonitrile (Table 1, entry **1k**). However, chloroform proved to be a useful alternative solvent for the iodination of **8** to afford **8a** in 83% yield, while the same reaction in acetonitrile gave only a 74% yield. This enhancement was not observed with 2-chloro and 2-bromophenol (**7** and **9**). Acetonitrile gave the highest yields of **7a** and **9a**. Nevertheless, chloroform is an alternative solvent for clean regioselective iodination in some cases (Table 1, entries **1k** and **8d**).

Lowering the temperature to 0 °C did not have a significant impact on the regioselectivity of phenol iodination. The selectivity of **1a** was 94% (Table 1, entry **1e**) after 8 h at 0 °C, while the selectivity of **1a** after 8 h at room temperature was 93% (Table 1, entry **1d**). Thus, subsequent monoiodinations of other substrates (compounds **2–11**) were conducted at room temperature using 1 equiv each of *p*TsOH and NIS. Iodination of *ortho*-substituted phenols occurred almost exclusively at the *para* position (relative to the OH). For all cases, except for **3**, which showed an amount of less than 1% for iodination at position 6, iodinations at position 6 of other substrates were no higher than 4%. For the *para*-substituted phenols, monoiodination occurred almost exclusively. For both *para* and *ortho*-substituted phenols, including substrate **1** also, monoiodination was the norm.

A preliminary study of bromination using *p*TsOH and *N*-bromosuccinimide (NBS) showed that bromination of phenol occurred

almost exclusively at the *para* position with a yield of 92%. The versatility of the iodination methodology is further demonstrated in the one-pot synthesis of 2-bromo-6-chloro-4-iodophenol (**7b**) via iodination and bromination. First, using 1.15 equiv of NIS (14 h) followed by 1.2 equiv of NBS (4 h) at room temperature, **7** was converted into **7b** in 74% yield (Table 1, entry **7e**).<sup>15</sup> Studies are underway to further explore the application of this methodology to monobromination and the one-pot sequential mixed halogenations of phenol analogues, and the results will be reported in due course.

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8. **Compound 1a**: mp 93 °C (lit.<sup>7a</sup> mp 93–94 °C); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 5.10 (br s, 1H, OH), 6.62 (d, *J* = 9 Hz, 2H), 7.51 (d, *J* = 9 Hz, 2H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 82.8, 117.8, 138.4, 155.2; GC/MS (EI) *m/z* (rel int.): 220 (100, M<sup>+</sup>), 191 (2, (M-CHO)<sup>+</sup>), 127 (4, 1), 110 (4), 93 (26, (M-1)<sup>+</sup>), 74 (2), 65 (23), 53 (3), 39 (10).
9. **Compound 2a**: colourless oil (lit.<sup>7b</sup> light-brown oil); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 2.24 (s, 3H), 5.17 (br s, 1H, OH), 6.87 (d, *J* = 8 Hz, 1H), 7.03 (d, *J* = 8 Hz, 1H), 7.47 (s, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 19.9, 85.4, 114.7, 130.9, 131.9, 138.3, 152.6; GC/MS (EI) *m/z* (rel int.): 234 (100, M<sup>+</sup>), 127 (4, 1), 107 (33, (M-1)<sup>+</sup>), 77 (28), 63 (3), 51 (9), 39 (3).
10. **Compound 3a**: mp 65–67 °C (lit.<sup>7c</sup> mp 66.5–68 °C); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 2.20 (s, 3H), 4.79 (br s, 1H, OH), 6.54 (d, *J* = 8 Hz, 1H), 7.35 (dd, *J* = 2 and 8 Hz, 1H), 7.43 (d, *J* = 2 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 15.4, 82.6, 117.1, 126.7, 135.8, 139.5, 153.7; GC/MS (EI) *m/z* (rel int.): 234 (100, M<sup>+</sup>), 127 (4, 1), 107 (20, (M-1)<sup>+</sup>), 77 (34), 63 (4), 51 (7), 39 (4).
11. **Compound 4a**: colourless oil (lit.<sup>7d</sup> mp 43 °C); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 3.87 (s, 3H), 5.59 (s, 1H, OH), 6.68 (d, *J* = 8 Hz, 1H), 7.18 (dd, *J* = 2 and 8 Hz, 1H), 7.22 (d, *J* = 2 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 56.2, 80.9, 116.5, 119.7, 130.4, 145.7, 147.4; GC/MS (EI) *m/z* (rel int.): 251 (8, (M+1)<sup>+</sup>), 250 (100, M<sup>+</sup>), 236 (3, ((M+1)-CH<sub>3</sub>)<sup>+</sup>), 235 (45, (M-CH<sub>3</sub>)<sup>+</sup>), 208 (1, ((M+1)-CH<sub>3</sub>-CO)<sup>+</sup>), 207 (20, (M-CH<sub>3</sub>-CO)<sup>+</sup>), 123 (4, (M-1)<sup>+</sup>), 108 (7, (M-1-CH<sub>3</sub>)<sup>+</sup>), 94 (2), 80 (5), 79 (5), 63 (5), 52 (11).
12. **Compound 5a**: mp 90 °C (lit.<sup>7e</sup> mp 91–92 °C); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 2.64 (s, 3H), 6.78 (d, *J* = 9 Hz, 1H), 7.71 (dd, *J* = 2 and 9 Hz, 1H), 8.00 (d, *J* = 2 Hz, 1H), 12.18 (s, 1H, OH); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 26.7, 79.7, 120.9, 139.2, 144.7, 152.7, 161.9, 203.5; GC/MS (EI) *m/z* (rel int.): 262 (100, M<sup>+</sup>), 247 (77, (M-CH<sub>3</sub>)<sup>+</sup>), 219 (14, (M-COCH<sub>3</sub>)<sup>+</sup>), 191 (4), 135 (3, (M-1)<sup>+</sup>), 127 (3, 1), 120 (7), 107 (3), 92 (12), 77 (5), 63 (12), 43 (12).
13. **Compound 6a**: mp 62–63 °C (lit.<sup>7f</sup> mp 78 °C); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 3.96 (s, 3H), 6.77 (d, *J* = 9 Hz, 1H), 7.69 (dd, *J* = 2 and 9 Hz, 1H), 8.13 (d, *J* = 2 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 52.6, 80.1, 114.5, 120.0, 138.3, 144.1, 161.2, 169.4; GC/MS (EI) *m/z* (rel int.): 279 (7, (M+1)<sup>+</sup>), 278 (85, M<sup>+</sup>), 247 (17, ((M+1)-CH<sub>3</sub>O-H)<sup>+</sup>), 246 (100, (M-CH<sub>3</sub>O-H)<sup>+</sup>), 219 (6), 218 (32), 191 (3), 190 (4), 151 (3, (M-1)<sup>+</sup>), 127 (3, 1), 120 (3), 119 (3), 92 (8), 91 (9), 63 (22), 53 (8), 38 (2).
14. **Compound 7a**: mp 53–54 °C (lit.<sup>7g</sup> mp 54 °C); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 5.57 (br s, 1H, OH), 6.78 (d, *J* = 9 Hz, 1H), 7.46 (dd, *J* = 2 and 9 Hz, 1H), 7.62 (d, *J* = 2 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 81.5, 118.2, 125.0, 136.9, 137.3, 151.4; GC/MS (EI) *m/z* (rel int.): 256 (32, (M+2)<sup>+</sup>), 254 (100, M<sup>+</sup>), 129 (8, ((M+2)-1)<sup>+</sup>), 127 (34, (M-1)<sup>+</sup>), 99 (21), 91 (10), 73 (8), 63 (21).
15. **Compound 7b**: mp 82–84 °C (lit.<sup>7h</sup> mp 81–82 °C); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 5.99 (s, 1H, OH), 7.46 (d, *J* = 2 Hz, 1H), 7.74 (d, *J* = 2 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 83.9, 112.9, 119.7, 132.0, 139.6, 150.4; GC/MS (EI) *m/z* (rel int.): 336 (26, (M+4)<sup>+</sup>), 334 (100, (M+2)<sup>+</sup>), 332 (78, M<sup>+</sup>), 305 (1, ((M+2)-CHO)<sup>+</sup>), 303 (1, (M-CHO)<sup>+</sup>), 209 (1, ((M+4)-1)<sup>+</sup>), 207 (3, ((M+2)-1)<sup>+</sup>), 205 (2, (M-1)<sup>+</sup>), 181 (6), 179 (26), 177 (19), 143 (6), 141 (7), 127 (9, 1), 97 (13, (M-Br-1-CHO)<sup>+</sup>), 62 (20, (M-Br-Cl-1-CHO)<sup>+</sup>), 53 (5).
16. **Compound 8a**: mp 75–76 °C (lit.<sup>7i</sup> mp 75–77 °C); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 5.34 (s, 1H, OH), 6.91 (d, *J* = 9 Hz, 1H), 7.21 (dd, *J* = 2 and 9 Hz, 1H), 7.63 (d, *J* = 2 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 85.4, 115.7, 126.1, 130.1, 137.2, 153.8; GC/MS (EI) *m/z* (rel int.): 256 (33, (M+2)<sup>+</sup>), 254 (100, M<sup>+</sup>), 225 (1, (M-CHO)<sup>+</sup>), 129 (4, ((M+2)-1)<sup>+</sup>), 127 (19, (M-1)<sup>+</sup>), 101 (7), 99 (23), 73 (7), 63 (15), 53 (4).
17. **Compound 9a**: mp 49–50 °C (lit.<sup>7j</sup> mp 51 °C); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 5.49 (s, 1H, OH), 6.71 (d, *J* = 9 Hz, 1H), 7.41 (dd, *J* = 2 and 9 Hz, 1H), 7.67 (d, *J* = 2 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 81.0, 110.3, 117.1, 137.0, 138.6, 151.3; GC/MS (EI) *m/z* (rel int.): 300 (98, (M+2)<sup>+</sup>), 298 (100, M<sup>+</sup>), 271 (1, ((M+2)-CHO)<sup>+</sup>), 269 (1, (M-CHO)<sup>+</sup>), 173 (16, ((M+2)-1)<sup>+</sup>), 171 (17, (M-1)<sup>+</sup>), 145 (11), 143 (12), 127 (7, 1), 119 (3), 117 (3), 92 (19), 74 (3), 63 (28), 53 (8), 38 (3).
18. **Compound 10a**: mp 92–93 °C (lit.<sup>7j</sup> mp 91–92 °C); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 7.25 (d, *J* = 8 Hz, 1H), 7.38 (t, *J* = 8 Hz, 1H), 7.55 (t, *J* = 8 Hz, 1H), 7.74 (d, *J* = 8 Hz, 2H), 7.93 (d, *J* = 8 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 86.2, 116.4, 124.2, 128.2, 128.3, 129.7, 130.3, 130.6, 134.8, 153.8; GC/MS (EI) *m/z* (rel int.): 270 (100, M<sup>+</sup>), 241 (2, (M-CHO)<sup>+</sup>), 143 (9, (M-1)<sup>+</sup>), 127 (2, 1), 115 (36), 89 (4), 74 (2), 63 (5) 39 (1).
19. **Compound 11a**: mp 78 °C (dec) (lit.<sup>20a</sup> mp 104–105 °C; lit.<sup>20b</sup> mp 104 °C); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 5.48 (br s, 1H, OH), 6.64 (d, *J* = 8 Hz, 1H), 7.55 (dt, *J* = 2 and 8 Hz, 1H), 7.62 (dt, *J* = 2 and 8 Hz, 1H), 7.89 (d, *J* = 8 Hz, 1H), 8.06 (dd, *J* = 1 and 8 Hz, 1H), 8.19 (dd, *J* = 1 and 8 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 88.2, 108.2, 110.1, 122.3, 126.1, 128.3, 131.9, 135.1, 136.8, 152.4; GC/MS (EI) *m/z* (rel int.): 270 (100, M<sup>+</sup>), 241 (3, (M-CHO)<sup>+</sup>), 207 (1), 144 (10, ((M+1)-1)<sup>+</sup>), 143 (11, (M-1)<sup>+</sup>), 135 (5), 127 (3, 1), 115 (52), 89 (7), 63 (7), 32 (5).
20. (a) Sket, B.; Zupet, P.; Zupan, M. *J. Chem. Soc., Perkin Trans. 1* **1989**, 2279–2281; (b) Sumi Mitra, S.; Sreekumar, K. *React. Funct. Polym.* **1997**, *32*, 281–291. At this stage we are unable to explain the discrepancy between our melting point and the literature melting points. The <sup>1</sup>H NMR and MS data of **11a** matched those reported in the literature.